AN ANALYTICAL NEW RP-HPLC METHOD FOR THE QUANTITATIVE DETERMINATION OF MOLNUPIRAVIR IN BULK AND TABLET DOSAGE FORM

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ABSTRACT: A rapid and precise reverse phase high performance liquid chromatographic method has been developed for the validated of Molnupiravir, in its pure form as well as in pharmaceutical dosage form. Chromatography was carried out on a Phenomenex Luna C18, 150 mm x 4.6 mm and 5 μ m column using a mixture of Methanol and Phosphate Buffer was in the ratio of 25:75%v/v (pH-3.4) as the mobile phase at a flow rate of 1.0ml/min, the detection was carried out at 270 nm. The retention time of the Molnupiravir was found to be 2.784 ±0.02min respectively. The method produce linear responses in the concentration range of 5-15 μ g/ml of Molnupiravir. The method precision for the determination of assay was below 2.0 %RSD. The method is useful in the quality control of bulk and pharmaceutical formulations.

Keywords: Molnupiravir, RP-HPLC, Method Development, Validation, Accuracy, Precision.

I. INTRODUCTION

Molnupiravir is an orally bioavailable prodrug of EIDD-1931, the synthetic ribonucleoside derivative N4hydroxycytidine and ribonucleoside analog, with potential antiviral activity against a variety of RNA viruses. Upon oral administration, Molnupiravir¹, being a prodrug, is metabolized into its active form EIDD-1931 and converted into its triphosphate (TP) form. The TP form of EIDD-1931 is incorporated into RNA and inhibits the action of viral RNA-dependent RNA polymerase. This results in the termination of RNA transcription and decreases viral RNA production, and viral RNA replication. Molnupiravir is hydrolyzed in vivo to N4hydroxycytidine, which is phosphorylated in tissue to the active 5'-triphosphate form, and incorporated into the genome of new virions, resulting in the accumulation of inactivating mutations, known as viral error catastrophe. A Remdesivir resistant mutant mouse hepatitis virus has also been shown to have increased sensitivity to N4hydroxycytidine. Molnupiravir² is indicated for treatment of mild to moderate coronavirus disease (COVID-19) in adults with a positive SARS-COV-2 diagnostic test and who have at least one risk factor for developing severe illness. The IUPAC Name of Molnupiravir³ is [(2R, 3S, 4R, 5R)-3, 4-dihydroxy-5-[4-(hydroxy amino)-2oxopyrimidin-1-yl] oxolan-2-yl] methyl 2-methyl propanoate. The Chemical Structure of Molnupiravir is as following



Fig.1. Chemical Structure of Molnupiravir

S No	Instruments/Equipments/Apparatus
D •1 10 •	mstruments/ Equipments/ Apparatus
1.	HPLC WATERS with Empower2 Software with Isocratic with UV-Visible
	Detector.
2.	T60-LABINDIA UV – Vis spectrophotometer
3.	High Precision Electronic Balance
4.	Ultra Sonicator (Wensar wuc-2L)
5	Thermal Oven
5.	Therman Oven
6.	Symmetry C_{18} Column, 250 mm x 4.6 mm and 5µm particle size
7	D ^H Analyzer (ELICO)
7.	F Allalyzer (ELICO)
8.	Vaccum Filtration Kit (Labindia)

II. MATERIALS AND METHODS Table 1. List of Fauinmonts

Table-2: List of Chemicals used

S.No.	Name	Grade	Manufacturer/Supplier
1	HPLC grade water	НРІ С	Sd fine-Chem ltd: Mumbai
1.			Su file chem ha, Mullou
2.	Methanol	HPLC	Loba Chem; Mumbai.
3.	Ethanol	A.R.	Sd fine-Chem ltd; Mumbai
4.	Acetonitrile	HPLC	Loba Chem; Mumbai.
5.	DMSO	A.R.	Sd fine-Chem ltd; Mumbai
6.	DMF	A.R.	Sd fine-Chem ltd; Mumbai

HPLC Instrumentation & Conditions:

The HPLC system employed was HPLC WATERS with Empower2 Software with Isocratic with UV-Visible Detector⁴.

Standard preparation for UV-spectrophotometer analysis:

The standard stock solutions - 10 mg of Molnupiravir standard was transferred into 10 ml volumetric flask, dissolved & make up to volume with Methanol. Further dilutions were done by transferring 1 ml of the above solution into a 10ml volumetric flask and make up to volume with methanol to get 10ppm concentration.

It scanned in the UV spectrum in the range of 200 to 400nm. This has been performed to know the maxima of Molnupiravir, so that the same wave number can be utilized in HPLC UV detector for estimating the Molnupiravir.

Different Trials for Chromatographic Conditions:

Table-3: Different Chromatographic Conditions								
Mobile Phase	Flow Rate	Wave length	Observation	Result				
	Table-3: Different Ch Mobile Phase	Table-3: Different Chromatographic Mobile Phase Flow Rate	Table-3: Different Chromatographic Conditions Mobile Phase Flow Rate Wave length Interview Interview	Table-3: Different Chromatographic Conditions Mobile Phase Flow Rate Wave Observation length length length length				

Inertsil C ₁₈ , 250 mm x 4.6 mm and 5µm Column	Acetonitrile : Water (60:40)	0.8 ml/min	270nm	Broad Peak	Method rejected
Symmetry C_{18} , 150 mm x 4.6 mm and 5 μ m Column	Methanol : Water (70:30)	0.9 ml/min	270nm	Negligible Peaks	Method rejected
Hypersil C ₁₈ , 250 mm x 4.6 mm and 5µm Column	Methanol : Acetonitrile (80:20)	0.9 ml/min	270nm	Multiple Peaks	Method rejected
Develosil C ₁₈ , 150 mm x 4.6 mm and 5µm Column	Methanol : Acetonitrile (70:30)	1.0 ml/min	270nm	Base line noise is high	Method rejected
Zorbax C ₁₈ , 250 mm x 4.6 mm and 5µm Column	Methanol : Acetonitrile (40:60)	1.0 ml/min	270nm	Stabilization not done properly	Method rejected
Phenomenex Luna C ₁₈ , 150 mm x 4.6 mm and 5µm Column	Methanol : Phosphate Buffer (45:55) (pH-4.2)	1.0 ml/min	270nm	Extra peaks	Method rejected
Phenomenex Luna C ₁₈ , 150 mm x 4.6 mm and 5µm Column	Methanol : Phosphate Buffer (35:65) (pH-3.8)	1.0 ml/min	270nm	Extra peaks & stabilization was not done properly	Method rejected
Phenomenex Luna C ₁₈ , 150 mm x 4.6 mm and 5µm Column	Methanol : Phosphate Buffer (25:75) (pH-3.4)	1.0 ml/min	270nm	Good sharp peak	Method accepted

Preparation of Phosphate buffer pH-3.4:

Accurately weighed 6.8 grams of KH2PO4 was taken in a 1000ml volumetric flask, dissolved and diluted to 1000ml with HPLC water and the pH was adjusted to 3.4 with orthophosphoric acid solution.

Preparation of Mobile Phase:

The mobile phase used in this analysis containing of a mixture of Methanol and Phosphate Buffer in the ratio of 25:75% v/v was prepared in the volume of 1000ml in which 250ml of Methanol was mixed with 750ml of Water. **Preparation of Standard Solution:**

Working concentration should be about $10\mu g/ml$. Correctly weigh around 10mg of Molnupiravir working standard, poured into a clean and dry 10 ml volumetric flask. Then dissolved and diluted to volume with the mobile phase to obtain a solution having a known concentration of about 1000 mcg/ml or 1000ppm. Further dilutions have been made to get the final concentration of $10\mu g/ml$.

III. RESULTS AND DISCUSSION Method Development





Observation: While scanning the Molnupiravir solution we observed the maxima at 270nm. **Optimized Chromatographic Conditions:**

Table-4:	Optimized	Chromatographic	Conditions
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Column	Phenomenex Luna C_{18} , 150 mm x 4.6 mm and 5µm
Mobile Phase	Methanol : Phosphate Buffer (25:75) (pH-3.4)
Flow Rate	1.0ml/minute
Wave length	270 nm
Injection volume	10 µl
Run time	7 minutes
Column Temperature	Ambient



Fig.3. Optimized Chromatogram for Molnupiravir

Method Validation

Table-5.

System Suitability Test System suitability testing⁵⁻⁷ is an integral part of many analytical procedures. The tests are based on the concept that the equipment, electronics, analytical operations and samples to be analysed constitute an integral system that can be evaluated as such. Following system suitability test parameters were established. The data are shown in

Table-5. Data of System Suitability Test								
S.No	Injection no	RT	Area	Height	USP Plate Count	USP Tailing		
1	Injection 1	2.768	2435648	7542	8745	1.38		

Table-5: Data of System Suitability Test

2	Injection 2	2.784	2452417	7659	8758	1.39
3	Injection 3	2.786	2453654	7582	8975	1.35
4	Injection 4	2.794	2454576	7635	8965	1.38
5	Injection 5	2.780	2455645	7659	8859	1.34
6	Injection 6	2.789		7698	8869	1.37
			2453524			
Mean						
			2450911			
S.D						
			7555.422			
%RSD						
			0.30827			

Accuracy:

Recovery study:

To determine the accuracy⁸ of the proposed method, recovery studies were carried out by adding different amounts (50%, 100%, and 150%) of pure drug of Molnupiravir were taken and 3 replications of each has been injected to HPLC system. From that percentage recovery values⁹ were calculated from the linearity equation y = 242891x + 28359. The results were shown in table-6.

Sample ID	Concentration (µg/ml)		Posk Aros	% Recovery of Pure drug	Statistical Analysis	
	Amount Injected	Amount Recovered		T ure urug	Analy 515	
S ₁ : 50 %	5	4.998	1242548	99.960%	Mean =	% Mean Recovery = 100 16%
S ₂ : 50 %	5	4.996	1241879	99.920%	100.22% S.D. =	100.1070
S ₃ : 50 %	5	5.039	1252436	100.780%	0.004854 % R.S.D.= 0.484321	
S ₄ : 100 %	10	9.998	2456854	99.980%	Mean = 100.05%	
S ₅ : 100 %	10	9.983	2453216	99.830%	S.D. = 0.002621	
S ₆ : 100 %	10	10.034	2465648	100.340%	% R.S.D. = 0.261976	
S ₇ : 150 %	15	15.016	3675847	100.106%	Mean = 100.20%	
S ₈ : 150 %	15	15.043	3682345	100.286%	S.D. = 0.000911	
S ₉ : 150 %	15	15.033	3679868	100.220%	% R.S.D. = 0.090875	

 Table-6: Accuracy Readings

Observation: From the Accuracy Method, we observed that the mean %Recovery of the drug are 100.22%, 100.05% and 100.20% which is within the range of 98-102% and %RSD is within the range <2, i.e. 0.484321%, 0.261976% and 0.090875% respectively. **Precision:**

Repeatability

The precision¹⁰ of each method was ascertained separately from the peak areas & retention times obtained by actual determination of six replicates of a fixed amount of drug Molnupiravir (API). The percent relative standard deviation¹¹ was calculated for Molnupiravir.

HPLC Injection	Retention	Peak Area	Theoretical	Tailing
Replicates of Molnupiravir	Time		Plates	Factor
Replicate – 1			8652	1.38
	2.777	2453658		
Replicate – 2			8694	1.34
	2.799	2469854		
Replicate – 3	2.789		8672	1.39
		2458677		
Replicate – 4	2.797		8692	1.37
		2459689		
Replicate – 5			8679	1.35
	2.797	2477898		
Replicate – 6			8638	1.36
	2.799	2469853		
Average				
		2464938		
Standard Deviation				
		9058.437		
% RSD				
		0.367491		

Table-7: Repeatability readings

Observation: From the Precision method, we observed that the %RSD of the Peak Area is 0.367491 which are within the acceptable range as per ICH guidelines¹²⁻¹⁵.

Intermediate precision:

The Intermediate Precision¹⁶ consists of two methods:-

Intra Day: In Intra Day process, the 50%, 100% and 150% concentration are injected at different intervals of time in same day.

Inter Day: In Inter Day process, the 50%, 100% and 150% concentration are injected at same intervals of time in different days.

Table-8: Results of Intermediate precision Day 1 for Monupiravir								
S.No.	Peak Name	RT	Area (µV*sec)	Height (µV)	USP Plate Count	USP Tailing		
1	Molnupiravir	2.768	2489851	7584	8658	1.38		
2	Molnupiravir	2.789	2478658	7598	8699	1.37		
3	Molnupiravir	2.784	2478785	7596	8693	1.39		
Mean			2482431					
Std. Dev.			6425.934					
% RSD			0.258856					

Intra-Day (Day 1): Table-8: Results of Intermediate precision Day 1 for Molnupiravir

Inter Day (Day 2):

-									
S.No.	Peak Name	RT	Area (µV*sec)	Height (µV)	USP Plate Count	USP Tailing			
1	Molnupiravir	2.777	2536542	7869	8987	1.57			
2	Molnupiravir	2.780	2547154	7889	8967	1.56			
3	Molnupiravir	2.797	2569853	7896	8969	1.59			
Mean									
Std. Dev.									
% RSD									

Table-9: Results of Intermediate precision Day 2 for Molnupiravir

Observations: The intra & inter day variation¹⁷ of the method was carried out for standard deviation & % RSD (% RSD <2%) within a day & day to day variations for Molnupiravir revealed that the proposed method is precise. **Linearity & Range:**

To evaluate the linearity, serial dilution of analyte were prepared from the stock solution was diluted with mobile phase¹⁸ to get a series of concentration ranging from $5-15\mu$ g/ml. The prepared solutions were sonicated. From these solutions, 10μ l injections of each concentration were injected into the HPLC system¹⁹ and chromatographed under the optimized conditions. Calibration curve²⁰ was constructed by plotting the mean peak area (Y-axis) against the concentration (X-axis).

Tuble 10. Concentration of Montuphavin					
Concentration in ppm	Peak Area				
0	0				
5	1275475				
7.5	1857648				
10	2456587				
12.5	3082546				
15	3642447				

Table-10: Concentration of Molnupiravir



Fig.4. Calibration Curve of Molnupiravir (API)

Observation: We observed that the calibration curve showed good linearity in the range of 5-15 μ g/ml, for Molnupiravir (API) with correlation coefficient (R²) of 0.9996. A typical calibration curve has the regression equation²¹ of y = 242891x + 28359 for Molnupiravir.

Method Robustness:

Influence of small changes in chromatographic conditions such as change in flow rate 1ml (\pm 0.1ml/min), Wavelength of detection 270nm (\pm 2nm) & organic phase content in mobile phase (\pm 5%) studied to determine the robustness²² of the method are also in favour of (Table-11, % RSD < 2%) the developed RP-HPLC method for the analysis of Molnupiravir (API).

Table-11: Kesuits of Method Kobustness Test					
Change in Parameter	Theoretical Plates	Tailing Factors	% RSD		
Flow (1.0 ml/min)	8645	1.38	0.265		
Flow (0.9 ml/min)	8754	1.37	0.487		
More Organic (75+5)	8952	1.32	0.698		
Less Organic (75-5)	8214	1.34	0.458		
Wavelength of Detection (272 nm)	8368	1.39	0.597		
Wavelength of detection (268nm)	8952	1.34	0.856		

Table-11: Results of Method Robustness Test

Limit of Detection (LOD) & Limit of Quantitation (LOQ):

The detection $limit^{23}$ (LOD) and quantization $limit^{24}$ (LOQ) may be expressed as:

$$L.O.D. = 3.3(SD/S).$$

$$L.O.Q. = 10(SD/S)$$

Where, SD = Standard deviation of the response

S = Slope of the calibration curve

The slope S may be estimated from the calibration curve of the analyte.

The Minimum concentration level at which the analyte can be reliable detected (LOD) & quantified (LOQ) were found to be $0.6902\mu g/ml \& 2.091\mu g/ml$ respectively.

Estimation of Molnupiravir in TABLET Dosage Form

Twenty Capsules were taken and the I.P. method was followed to determine the average weight. Above weighed Capsules were finally powdered and triturated well. A quantity of powder equivalent to 10 mg of drug were transferred to 10 ml volumetric flask, and 8 ml of mobile phase was added and solution was sonicated for 15 minutes, there after volume was made up to 10 ml with same solvent. Then 1ml of the above solution was diluted to 10 ml with HPLC grade methanol. The solution was filtered through a membrane filter (0.45 μ m) and sonicated to degas. From this stock solution (1.0 ml) was transferred to five different 10 ml volumetric flasks and volume was made up to 10 ml with same solvent system.

The solution prepared was injected in five replicates into the HPLC system and the observations were recorded. A duplicate injection of the standard solution was also injected into the HPLC system and the peak areas were recorded. The data are shown in Table-12.

ASSAY

% Assay=AT/AS×WS/DS×DT/WT×P/100×AW/LC×100

Where:

AT = Peak Area of Molnupiravir obtained with test preparation

AS = Peak Area of Molnupiravir obtained with standard preparation

WS = Weight of working standard taken in mg

WT = Weight of sample taken in mg

DS = Dilution of Standard solution

DT = Dilution of sample solution

P = Percentage purity of working standard

Brand Name of Capsules	Labelled amount of Drug (mg)	Mean (±SD) amount (mg) found by the proposed method (n=5)	Assay + % RSD
Molflu Capsules (Dr. Reddy's)	200	199.867 (± 0.324)	99.867% (± 0.457)

Results obtained are tabulated below: Table-12: Assay of Molnupiravir Capsule

Result & Discussion: The %Purity²⁵ of Molflu Capsule containing Molnupiravir was found to be 99.867% (\pm 0.457).

Stability Studies:

Results of Degradation Studies: The results of the stress studies²⁶⁻³⁰ indicated the specificity of the method that has been developed. Molnupiravir was stable in thermal, Oxidation and Photolytic stress conditions. The results of forced degradation studies are given in the following table-13.

Stress Condition	Time	Assay of active substance	Assay of degraded products	Mass Balance (%)
Acid Hydrolysis (0.1N HCl)	24Hrs.	76.302	23.698	100%
Basic Hydrolysis (0.1N NaOH)	24Hrs.	84.426	15.574	100%
Thermal Degradation (60 ⁰ C)	24Hrs.	91.105	8.895	100%
UV (254nm)	24Hrs.	85.121	14.879	100%
3% Hydrogen peroxide	24Hrs.	86.315	13.685	100%

Table-13: Results of Forced Degradation Studies of Molnupiravir

IV. SUMMARY AND CONCLUSION

The analytical method was developed by studying different parameters. First of all, maximum absorbance was found to be at 270 nm and the peak purity was excellent. Injection volume was selected to be 10μ l which gave a good peak area. The column used for study was Phenomenex Luna C18, 150 mm x 4.6 mm and 5μ m because it was giving good peak. Ambient temperature was found to be suitable for the nature of drug solution. The flow rate was fixed at 1.0ml/min because of good peak area and satisfactory retention time.

Mobile phase is Methanol and Phosphate Buffer was taken in the ratio of 25:75% v/v at pH-3.4 was fixed due to good symmetrical peak. So this mobile phase was used for the proposed study. Run time was selected to be 7 min because analyze gave peak around 2.784 ± 0.02 min respectively and also to reduce the total run time. The percent recovery was found to be 98.0-102 was linear and precise over the same range. Both system and method precision was found to be accurate and well within range.

The analytical method was found linearity over the range $5-15\mu g/ml$ of Molnupiravir of the target concentration. The analytical passed both robustness and ruggedness tests. On both cases, relative standard deviation was well satisfactory.

REFERENCES

- [1] https://go.drugbank.com/drugs/DB15661
- [2] https://pubchem.ncbi.nlm.nih.gov/compound/eidd-2801
- [3] https://en.wikipedia.org/wiki/Molnupiravir
- [4] Journal of Pharmaceutical and Biomedical Analysis Volume 21, Issue 2, 1 November 1999, Pages 371–382.
- [5] Tropical Journal of Pharmaceutical Research, October 2009; 8 (5): 449-454 © Pharmacotherapy Group.
- [6] Instrumental Method of Analysis by Rabi Sankar, P-18-6, P-18-3.
- [7] Practical HPLC Method Development by Lloyd R. Snyder *et al*; 2nd edition, P-503.

- [8] Guidance for industry, Analytical Procedure and Method Validation, U.S. Department of Health and Human Services FDA, August 2000.
- [9] Y. F. Cheng, T.H. Walter, Z. Lu, P. Iraneta, C. Gendreau, U. D. Neue, J. M. Grassi, J. L. Carmody, J. E. O' Gara, and R. P. Fisk, LCGC 18(10), 1162 (2000).
- [10] The United State Pharmacopeia 25/National Formulary 20, ch. 1225, pg. 2256-2259 (The United State Pharmacopoeial Convention, Inc., Rockville, Maryland, 2002).
- [11] ICH Q2B: Validation of Analytical Procedure; Methodology (International Conferences on Harmonization of Technical requirements for the registration of Drugs for Human use, Geneva, Switzerland, May 1997).
- [12] ICH Q2B: Validation of Analytical Procedure; Methodology (International Conferences on Harmonization of Technical requirements for the registration of Drugs for Human use, Geneva, Switzerland, Nov 2003).
- [13] M. V. Gorenstein, J. B. Li, J. Van Antwerp, and D. Chapman, LCGC 12(10), 768-772 (1994).
- [14] 16. P. M. Young and M. V. Gorenstein, LCGC 12(11), 832-838 (1994).
- [15] W. J. Warren, W. A. Stanick, M. V. Gorenstein, and P. M. Young, Bio techniques (2), 282-297 (1995).
- [16] Validation of Analytical Procedures, Methodology, and ICH Harmonized Tripartite Guideline, 108, 1996.
- [17] Mcclellan KJ & Plosker GI, Drugs; 58,143-157, (July 1999)
- [18] The complete drug reference; Martindale, Pharmaceutical Press 32 edition; 12th pg.
- [19] Watnabe, M. et al. Synthesis and Biological Activity of Methane Sulfonamide Pyramiding and N-Methane Sulfonyl Pyrrole-Substituted 3, 5-Dihydroxy-Heptonates, A Novel Series of HMG-Co A Reductase Inhibitors. Bioorg. Med. Chem. 5, 437-444, (1997).
- [20] FDA Drug Approvals List [Online] (Cited 26 Aug. 2003).
- [21] International conference on harmonization, "Q2a: text on validation of analytical procedures," federal register 60(40), 11260–11262 (1995).
- [22] International conference on harmonization, "q2b: validation of analytical procedures: methodology; availability," federal register 62(96), 27463–27467 (1997).
- [23] FDA, "analytical procedures and methods validation: chemistry, manufacturing and controls documentation; availability," federal register (notices) 65(169), 52776–52777 (2000).
- [24] G.A. Shabir, "Validation of HPLC Chromatography Methods for Pharmaceutical Analysis, Understanding the differences and similarities between validation requirements of FDA, the US pharmacopeia and the ICH," j. Chromatogr. A. 987(1-2), 57-66 (2003).
- [25] J. M. Green, a practical guide to analytical method validation, anal. Chem. News & features, 1 May 1996, pp. 305a–309a.
- [26] P. A. Winslow and r. F. Meyer, defining a master plan for the validation of analytical methods, j. Validation technology, pp. 361–367 (1997).
- [27] Aoac peer-verified methods program, manual on policies and procedures, arlington, va., USA (1998).
- [28] Citac/Eurachem, working group, international guide to quality in analytical chemistry: an aid to accreditation, (2002).
- [29] Journal of Chromatography .B, Analytical Technologies in the Biomedical and life Sciences. 2008 March 1; 863(2): 258-265. Published on 2008 Jan 18.
- [30] Matheson A.J., Noble S., Drugs, Volume 59, Number 4, April 2000, pp. 829-835(7).
- [31] Tuba Reçber, Selin Seda Timur, Sevilay Erdoğan Kablan, Fatma Yalçın, Tutku Ceren Karabulut, R. Neslihan Gürsoy, Hakan Eroğlu, Sedef Kır, Emirhan Nemutlu, A stability indicating RP-HPLC method for determination of the COVID-19 drug Molnupiravir applied using Nano formulations in permeability studies, Journal of Pharmaceutical and Biomedical Analysis, Volume 214, 30 May 2022, 114693.
- [32] Santhosh Illendula, Naveen Kumar Singhal; A Review: Novel analytical method development & validation for the determination of selected anti-cancer & anti-viral drugs, WJPPS 11(07) 2022, 533-566.
- [33] Santhosh Illendula, M Divya, Rajeswar Dutt, KNV Rao; RP HPLC method development & validation for forced degradation studies for the simultaneous estimation of abacavir & lamivudine in pure form & marketed formulation, Int. J of research 08(5) May 2019, 3342-3364.
- [34] Jahnavi Bandla* and S. Ganapaty, Development and Validation of a Stability-indicating Method for the Simultaneous Estimation of Sofosbuvir and Ledipasvir by RP-HPLC, Indian J Pharm Sci 2018; 80(6):1170-1176.